

## Solid Tumour Test Request Form Colorectal

Fill in patient details below or affix addressograph (top left) and send to Cell Pathology for sample preparation						
Patient Forename:				Clinician (address report to):		
Patient Surname:	urname:			Requested by:		
DoB:	NHS numb		mber:	Hospital N	lame ( <u>essential for report</u> )	
Sex:	Hospital N		oncologis (NHS Wa		Email Addresses (for reports): oncologists/pathologists/MDT coordinators ) (NHS Wales or NHS.net)	
Address:	Alternat	Alternative Hospital no:				
		-				
Postcode:		Date red	Date requested:			
Please note: Gene analysis relies on sampling <u>tumour tissue.</u>						
Tissue blocks for genomic analysis can no longer be accepted.						
This section is for completion by Pathology Laboratory   Pathologist: Pathology Hospital: Block Number:						
					block Number.	
Sampling method, biopsy type and fixation method.			Date sample ser	nt to AWMGS	Tumour sample has now been exhausted	
					Yes D No D	
For <u>ALL</u> requests please provide: <b>1</b> H&E stained slide with area of highest neoplastic cell content <u>CLEARLY circled</u> . Please state the approx. % neoplastic cell content present in the H&E circled tumour area:%						
Relevant Clinical Summary (e.g. tumour histology) Please also attach appropriate pathology report						
Test Test direct		Test directory	Technology	Sample requ	Sample requirements	
Multi-target DNA NGS panel: small variant - KRAS, NRAS, BRAF, TP53, PTEN, EGFR, PIK3CA		M1.1	DNA NGS Panel DNA: 60μM (preferably 6 x 10μM) air dried unstained sections mounted on slides.			
Multi-target RNA NGS panel: structural variant – NTRK1, NTRK2,		M1.6	RNA NGS Panel	sections mour prepared in ar	<b>RNA:</b> 50uM (preferably <b>5 x 10μM</b> ) air dried unstained ections mounted on slides. <i>Note: slides for RNA – ideally repared in an RNase-free environment.</i>	
NTRK3				cannot be per	SH (in the event that RNA-based NGS formed or is unsuccessful): 2 x 3-4μM y mounted) on charged/adhesion slides	