

Acute Myeloid Leukemia (AML) Service

Background

Acute myeloid leukemia (AML) is a genetically heterogeneous hematological malignancy of myeloid lineage, characterized by clonal expansion of myeloid blast cells.

There are many different classes of AML and several related neoplasms which are genetically defined within the WHO guidelines (2016). The genetic drivers affect many different genes and range from single nucleotide polymorphisms (SNPs) to larger chromosomal changes. A range of cytogenetic and molecular tests are routinely offered at AWMGL for the detection and monitoring of genetic abnormalities in patients with a diagnosis of AML. Next generation sequencing (NGS) is also available upon request.

Acute promyelocytic leukemia (APL) is a clinically and genetically distinct class of leukemia that is clinically urgent due to its rapid progression to disseminated intravascular coagulation (DIC). It is characterized by the WHO through a reciprocal translocation which leads to a fusion gene by juxtaposition of the PML gene (on chromosome 15) and the RARA gene (on chromosome 17). The translocation is present in 95-98% of cases (Rashidi and Fisher, 2015) and is a diagnostic and prognostic marker of APL, which can be targeted with all-*trans* retinoic acid (ATRA) therapy.

Test information

- **New or query AML cases will receive a diagnostic screen consisting of:**
 - G-band analysis for the detection of cytogenetic abnormalities.
 - Molecular analysis for the detection of
 - CFBF-MYH11 gene fusion transcripts A, D and E.
 - RUNX1-RUNX1T1 gene transcript.
 - FLT3-ITD mutation
 - FLT3 tyrosine kinase domain (FLT3-TKD) mutations
 - NPM1 mutations
- MRD testing by quantitative PCR is available for patients with a confirmed CFBF-MYH11/ RUNX1-RUNX1T1 gene fusion.
- New and query **APL cases** will be tested by rapid fluorescent in situ hybridisation (FISH) in addition to molecular testing for the PML-RARA fusion rearrangement. The PCR test is used for identification of three isoforms of the PML-RARA fusion transcript. The appropriate PCR test for that transcript may then be used for MRD testing to follow up response to treatment.
- **Next generation sequencing (NGS)** is available upon request for confirmed cases of AML.

Referral Criteria

All requests should be made on an appropriate request form available at the AWMGS website www.medicalgenomicswales.co.uk.

References

Arber, D.A., Orazi, A., Hasserjian, R., Thiele, J., Borowitz, M.J., Le Beau, M.M., Bloomfield, C.D., Cazzola, M. and Vardiman, J.W. (2016). The 2016 revision to the World Health Organization classification of myeloid

neoplasms and acute leukemia. *Blood*, [online] 127(20), pp.2391–405. Available at: <https://www.ncbi.nlm.nih.gov/pubmed/27069254/>.

Rashidi, A, and S I Fisher. "FISH-Negative, Cytogenetically Cryptic Acute Promyelocytic Leukemia." *Blood Cancer Journal*, vol. 5, no. 6, June 2015, pp. e320–e320, 10.1038/bcj.2015.47.

Contact Details

All Wales Genomics Laboratory,
Institute of Medical Genetics,
University Hospital of Wales,
Heath Park,
Cardiff CF14 4XW

Tel: 029 2074 2641

Fax: 029 2074 4043

Haematology.Genetics.cav@wales.nhs.uk

lab.genetics@wales.nhs.uk
www.wales.nhs.uk/AWMGS/
Accredited to ISO 15189:2012
(8988)

Sample Requirements

Bone marrow is preferred - send in sterile transport medium supplied by laboratory or in a lithium heparin blood tube.

Blood in lithium heparin for cytogenetic analysis

Blood in EDTA for molecular analysis

Please label samples with three identifiers and date of collection

All samples must be accompanied by a completed request form

Consent for testing and sample storage is assumed when the request is received – it is the responsibility of the referring clinician to ensure that appropriate consent has been obtained.

TAT (Calendar days)

G-band analysis	14CD
FISH for new APL	3CD
FLT3-ITD mutation	10 CD
FLT3-TKD & NPM1 mutation screens	14CD
Fusion transcripts screens	14 CD
NGS	21 CD